Research article

ANTIBACTERIAL, ANTIFUNGAL AND INSECTICIDAL ACTIVITIES OF THE CRUDE EXTRACT FRACTION OF BARK OF MILLETTIA OVALIFOLIA

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Abstract

The crude extract fraction of the of stem bark of *Millettia ovalifolia* was investigated for their antibacterial, antifungal and insecticidal activities. The results obtained showed that crude fraction showed no antibacterial activity, while in case of antifungal and insecticidal activities it showed 10% activity against *Microsporum canis* and 20% against *Rhyzopertha dominica, Tribolium castenum* and *Callosbruchuanalis* respectively. All these results showed that the crude fraction of this plant has good insecticidal activity and moderate antifungal activity, which in turn revealed that this plant is of high medicinal importance.

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Key words- Millettia ovalifolia bark, antibacterial, antifungal and insecticidal activities

INTRODUCTION

The genus Millettia belongs to family Leguminosae (Papilionaceae); consists of about 150 species distributed in the tropical and sub-tropical regions of the world. Only two species are present in Pakistan i.e. M. extensa and M. ovalifolia¹. M. ovalifolia locally known as shewa (in pashto), villayati shisham (in Urdu), and rose wood (in English)². It is a tall ornamental deciduous tree, and the leaves are imparipinnately compound while the fruits are cylindrical pods 6.5 to 9 cm long. The genus *Millettia* is known for the medicinal importance due to a rich source of variety of compounds. The major class of compounds of this genus is flavonoids. The medicinal and pharmacological properties of genus Millettia is highlighted below: The species of genus Millettia such as M. *conrauai* is well known for it's insecticidal, molluscicidal and pisticicidal activities². Some of the species; M. pachycarpa inhibited the activities of murine retroviral reverse transcriptase and human DNA polymerases³ Compounds; rotenone and 3α -hydroxyrotenone isolated from *M. pervilleana have* shown inhibition of TPAinduced ornithine decarboxylase at the level of its m RNA expression and recommended as promising cancer chemo-preventive agents. The prenylated isoflavanone, pervilleanone from M. pervilleana showed anticancer activity³, while *M. ovalifolia* contains hypotensive agents⁴. The flavonoids; chalcones isolated from *M. ovalifolia* showed anti-malarial activitiy⁵, and Isoflavonoids; griffonianone and maximalsoflavone isolated from M. griffoniana showed significant cytotoxicity ⁶. M. dura contains 6α , 12α -didehydro-6-oxodeguelin has good insecticidal activity and played a key role as a phytotoxin inhibitor of ornithine decarboxylase⁷. M. griffoniana contains griffonianone C which showed a potent estrogenic activity ⁶, and Millepurone ⁸ has antitumor promoted reported from M. atropurpurea. Osajin isolated from M. auriculata has anti-oxidant activity ⁹ while M. duchusnei contains rotenones used as potent insecticides. In some parts of Africa especially in Cameroon the plants of genus Millettia are used by different communities as a potent inhibitor of intestinal parasites in children's as well as colic besides oral treatment for boils ¹⁰.

Material and methods Experimental

Plant material

The bark of *M. o*valifolia was collected during the month of June 2008 from Pakistan Forest Institute Peshawar. The plant was identified and authenticated by Dr. Samin Jan, Associate Professor, Department of Botany, Islamia University, Peshawar, Pakistan. The voucher specimen (SJ-33) was deposited in the herbarium of Botany Department, Islamia University, Peshawar, Pakistan.

Extraction and isolation

The stem bark was shade dried and powdered (70 kg) were soaked in 5% aqueous methanol for one week (x3). The combined brownish extract was concentrated under reduce pressure by a vacuum rotary evaporator obtained brownish residue **F1** (5 kg) which was suspended in water and partitioned with *n*-hexane to get *n*-hexane fraction **FX** (1.2 kg). The *n*-hexane insoluble portion was acidified with HCl (pH 2) and subjected to further fractionation with ethyl acetate (x3), afforded ethyl acetate fraction **FX1** (1.0 kg), while remaining insoluble fraction was basified with ammonia (pH 8) and fractionated with chloroform; obtained chloroform fraction **FX2** (1.6 kg) and insoluble fraction **FX4** (0.8 kg) (**Scheme-26**). These fractions were studied for antifungal, antibacterial, phytotoxic and insecticidal activity. The chloroform fraction (500 g) was subjected to column chromatography using silica gel with *n*-hexane-chloroform increasing polarity to obtain several fractions, which were combined on the basis of TLC profile afforded major fraction which was subjected rechromatographed using silica gel and *n*-hexane-chloroform in increasing polarity furnished 56 fractions.

Antibacterial activity

The microorganisms; *B. subtilis ATCC 6633, E. coli ATCC 25922, Straptodirimus, S. flexenari (clinical isolate), S. aureus ATCC 25923, P. aeruginosa ATCC 27853 and S. typhi ATCC19430 were used for evaluation of antibacterial activity. The organisms were stored in muller hantin agar in the refrigerator at 4^{\circ}C prior to subculture. Antibacterial testing was carried out on the already developed agar well diffusion method to study the potency of the crude extract of <i>M. ovalifolia.* Broth media were prepared and the test organisms were transferred to the broth media from agar

plate and were grown at 37^{0} C for 24 hours. After 24 hours 25 ml of MHA were poured into each petri plate and cooled in sterile condition. The fresh culture was prepared from day old culture, after solidification of MHA in plate, 0.6 ml of fresh culture of test organism were poured on to MHA. Wells of 6 mm diameter were digged in to the medium by using sterile borer and 22 mg of crude extract fraction of the stem bark of *M. ovalifolia* was used against each organism. DMSO and standard antibiotic (Imipenum) were added into other wells. The plates were kept in sterilized inoculation chambers for 1 hour to facilitate diffusion of the antimicrobial agent into the medium. The plates were then incubated at 37 0 C for 24 hours and the diameters of the zone of inhibition of microbial growth were measured in millimeters.¹¹

Antifungal activity

The microorganisms; *T. longifusis, C. albicans ATCC 2091, A. flavus ATCC 32611, M. canis ATCC 11622, F. solani 11712* and *C. glaberata ATCC 2091* were used for antifungal assay. All these strains were maintained on agar slant at 4° C, the slant was allowed to activate at a temperature of 37 $^{\circ}$ C for duration of 3-4 days on nutrient agar (NA), for fungi, before any screening is made. The crude extract fraction was dissolved in DMSO (24 mg/ml) and sterile medium (5 ml) was placed in a test tube and inoculated with the sample solution (400μ g/ml) which was then kept in a slanting position at room temperature for overnight. The tubes were inoculated by a piece of fungus (4 mm diameter) from seven day old culture. The samples were then incubated for 7 days at 28°C and the fungal strain starts growth on the slant. The growth inhibition was observed and percentage growth inhibition was determined by calculating with reference to the positive control by applying the formula

100-linear growth and test (mm) % Inhibition =------ x 100 linear growth in control (mm)

Miconazole was used as standard drug a positive control.¹²⁻¹³

Results and discussion

1. Antibacterial activity of the crude fraction

The crude fraction showed no antibacterial activity against selected bacterial strains (Table-1).

Table-1: Antibacterial activity of the crude extract of M. ovalifolia stems bark

Micro	Gram	F1	DMSO	Imipenum
organisms			(-)	$10 \mu \text{g/Disc}$
				(+)
E. coli	-	-	-	35
B. subtilis	+	-	-	36
S. flexenari	-	-	-	36
S. aureus	+	-	-	43
S. aeruginosa	-	-	-	32
S. typhi	-	-	-	40
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Key words: Well size 6 mm, F1 = Crude fraction



Figure-1: Antibacterial activity of the crude fraction

Antifungal activity

The crude fraction of stem bark of *M. ovalifolia* was also investigated for antifungal bioassay against the selected fungal strains; *T. longifusis, C. albicans, A. flavus, M. canis, F. solani, C. glaberata* (Table-2). The crude extract (F1) showed 10 % bioactivity against *M. canis.*

Table-2: Antifungal activity of the crude extract of *M. ovalifolia* stems bark.



Figure-2: Antifungal activity of the crude extract fraction of *M. ovalifolia* stem bark

Fungal species	Standard (μ g/ml)	F1
Trichophyton	Miconazole 70.08	-
longifusis		
Candida albicans	Miconazole 110.8	-
Aspergilus flavus	Miconazole 20	-
Microsporum canis	Miconazole 98.4	10
Fusarium Solani	Miconazole 73.10	-
Candida glaberata.	Miconazole 110.8	-

Key words: F1 =Crude fraction

Insecticidal activity

The crude extract fraction (**Table- 3**; **Figure-3**) evaluated for insecticidal activity against the selected insects; *T. castaneum*, *R. dominica* and *C. bruchuanalis*. The results are summarized in table-3 which clearly indicated that the crude fraction has 20% activities against these selected insects.

Table- 3: Insecticidal activity of the crude fraction of *M. ovalifolia* stem bark.

Insect	NOs	F1
T. castaneum	100	20
R. dominica	100	20
C. huanalis	100	20

Key words: F1= Crude fraction



Figure-3: Insecticidal activity of the crude extract fraction of *M. ovalifolia* stem bark.

CASE STUDY

In the present study, an effort was made to scientifically verify bioassay of the crude fraction of stem bark of medicinal plant known as *M. ovalifolia* to further intricate its medicinal importance. The consequences obtained are given both in tabular and graphical form which reveals that it contain variety of therapeutic agents which further need to explore phytochemically.

CONCLUSIONS

From the results above it is clear that the medicinal plant *M. ovalifolia* possess high medicinal importance as clear from the bioassay results and in the current study. Further scientific approach is needed to explore phytochemically this plant.

REFERENCES

[1] Ali, S.I., (1977) Flora of Pakistan: Shamim Printing Press: Karachi, 100, 51.

[2] Abiy, Y., Solomon, Jacob, O. M., Hellen, A.O.R. (2003) Phytochemistry 64, 773.

[3] Abiy, Y.; Jacob, O. M.; Hellen, A. O. R. (2003) *Phytochemistry*, 64, 779.

[4] Caron, R. A.; Maran, J. M.; Montero, L.; Fernandozaigo; A.A., D. (1987) Plantes Medicanales et Phytotherapeutic, 21, 202.

[5] Choudhary, M. I.; Shawar, D. E.; Parveen, Z.; Jabbar, A.; I.; A.; Rahman, A.U. (1995) Phytochemistry, 40, 1243.

[6] Fuendjiep, V.; Nkengfack, A. E.; Fomum, Z. T.; Sondengam, B.L.; Bodo, B. (1998) Phytochemistry, 47, 113.

[7] Ito, C.; Masataka, I.; Hugh, T. W. T.; Harukuni, T.; Xiao, Y. M.; Teruo, M.; Takeshi, I.; Hoyoku, N.; Hiroshi, F. (2000) *Canc. Lett.*, *152*, 187

[8] Janaki, S.; Vijayasekaram, V. J. (1998) Biomedicine, 18, 86.

[9] Merz, K. W. G.; Schmidt. (1935) Arch. Pharm., 273, 1.

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[10] Ngamga, A., Fanso Free, S.N.Y., Tane, A., Fomum, Z.T. (2007) Fitoterapia 78, 276.

[11] NgadJui, B. T.; Abegaz, B. M.; Dufat, H.; Tillequin, F. O.. (2008) Phytochemistry, 69, 263.

[12] Palazzinoa, G.; Rasoanaivob, P.; Federicia, E.; Nicolettic, M.; Galeffia, C. (2003) Phytochemistry, 63, 474.

[13] Wolfrom, M. L.; Benton, F. L.; Gregory, A. S.; Hess, W. W.; Mahan, J. E.; Morgan, P. W. (**1939**) *J. Am. Chem. Soc.*, *61*, 2832.

[14] Yankep, E., Mbafor, J.T., Fomuma, Z.T., Steinbeck, C., Messanga, B.B., Nyasse, B., Budzikiewicz, H., Lenz, C., Schmickler, H. (**2001**) *Phytochemistry* 56, 363-368.